

Application	Amount	Volume ( $\mu\text{L}$ )	Conc. (ng/ $\mu\text{L}$ )	Quality	Size (bp)	Comment
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**Illumina Short-Read Sequencing**

<i>DNA Sequencing</i>	<p><b>General remarks and requirements for DNA samples:</b></p> <ul style="list-style-type: none"> <li>- Buffer: TRIS-HCL 10 mM or lowTE</li> <li>- 260/280 ratio 1.8-2.0 (or according to quality column)</li> <li>- In case of genome sequencing: high molecular for best results</li> </ul> <p><b>All samples of an order must be adjusted to a uniform concentration within the specifications. Samples that do not meet the requirements listed here cannot be processed and will be rejected!</b></p>					
<i>Whole Genome - Low input (incl. PCR)</i>	150 – 300ng	$\geq 15$	$> 10$	see general remarks above	-	FFPE possible with constraints in output quality
<i>Whole Genome - PCR Free</i>	$> 300$ ng	$\geq 15$	$> 20$	see general remarks above	-	
<i>Bacterial genome sequencing</i>	15 ng	$\geq 15$	1	see general remarks above		
<i>Whole Exome Sequencing</i>	$> 75$ ng	$\geq 15$	5	see general remarks above	-	FFPE possible if amount based on qPCR measurement is sufficient
<i>Gene Panel</i>	600 ng	$\geq 15$	40	260/280 ratio 1.7-2.2 260/230 ratio 1.2	-	-
<i>Amplicons</i>	$> 75$ ng	$\geq 15$	5	260/280 ratio $> 1.5$	-	less material possible on request
<i>Microbiome 16S</i>	150 ng	$\geq 15$	10	260/280 ratio $> 1.5$	-	less material possible on request
<i>Microbiome Shotgun Metagenomics</i>	75 ng	$\geq 15$	5	260/280 ratio $> 1.5$	-	less material possible on request

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## Illumina Short-Read Sequencing

### RNA Sequencing

**General remarks and requirements** for totalRNA samples:

- Buffer: nuclease free water
- DNase treated and cleaned up
- 260/280 ratio >2.0
- RQN  $\geq 8$ ; RQN  $\Delta$  between samples <1
- ultra low input (< 1 ng total amount) possible, but must be planned beforehand

**All samples of an order must be adjusted to a uniform concentration within the specifications.**

**Samples that do not meet the requirements listed here cannot be processed and will be rejected!**

<i>Whole Transcriptome – standard input</i>	>375 ng	$\geq 15$	25 - 80	see general remarks above	-	-
<i>Whole Transcriptome - low input</i>	>15 ng	$\geq 15$	1 - 5	see general remarks above	-	very low input amount may lead to constraints in output quality
<i>Expression Profiling (mRNA) – standard input</i>	>375 ng	$\geq 15$	25 - 80	see general remarks above	-	-
<i>Expression Profiling (mRNA) – low input</i>	>15 ng	$\geq 15$	1 - 5	see general remarks above	-	very low input amount may lead to constraints in output quality
<i>3'Prime RNA Seq -standard input</i>	>375 ng	$\geq 15$	25 - 80	see general remarks above	-	-
<i>3'Prime RNA Seq -low input</i>	>15 ng	$\geq 15$	1 - 5	see general remarks above	-	very low input amount may lead to constraints in output quality
<i>Small RNA Profiling (miRNA, lnc-RNA etc.)</i>	50 ng	$\geq 15$	50	see general remarks above	-	Low input possible on request

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<i>Sequencing of prepared NGS libraries</i>	50 ng	$\geq 15$	5	No primer dimer residuals	-	Buffer: TRIS-HCL 10 mM
<b>10X Single-Cell RNA-Sequencing</b>						
<i>Single-Cell Transcriptomics</i>	1.000 – 30.000 cells /sample	50 $\mu\text{l}$	500-1.000 cells/ $\mu\text{l}$	Vitality >70 %	-	-
<i>Spatial Transcriptomics</i>	Tissue sections placed on Visium slides, RNA extracted from tissue section RIN >7					
<b>Bionano Optical Mapping</b>						
<i>Bionano Saphyr Chip</i>	1 $\mu\text{g}$	-	>36	260/280 = 1.8 260/230 = 2.0-2.2	Megabase range	-

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<b>Pacific Biosciences Long-Read Sequencing</b>						
<i>Transcriptome (Iso-Seq)</i>	>2 $\mu\text{g}$	>15	>130	RIN 8-10 260/280 = 2.0 260/230 = 2.2	-	DNase digested, buffer: RNase free water
<b>Whole Genome Sequencing</b>						
<i>HiFi Reads – Standard Protocol</i>	>6 $\mu\text{g}$	>120	50	260/280 = 1.8 260/230 = 2.0-2.2	>50kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>HiFi Reads – Low Input</i>	>1 $\mu\text{g}$	>50	20	260/280 = 1.8 260/230 = 2.0-2.2	>50kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>HiFi Reads – Ultra-Low Input</i>	>30 ng	>15	2	-	>50kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>Continuous Long Reads</i>	>6 $\mu\text{g}$	>120	50	260/280 = 1.8 260/230 = 2.0-2.2	>50kb	homogenous HMW DNA, RNase digested, buffer: EB
<b>Amplicon Sequencing</b>						
	500 ng – 3 $\mu\text{g}$ (depends on size)	50	-	-	-	clean, target-specific, buffer: EB
<b>Multiplexed Microbial</b>						
	>1 $\mu\text{g}$ per sample	20-100	-	260/280 = 1.8 260/230 = 2.0-2.2	>20kb	-

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<b>Oxford Nanopore Long-Read Sequencing</b>						
<i>Transcriptome</i>						
<i>Direct mRNA Sequencing</i>	>100 ng polyA+ or 1 $\mu\text{g}$ Total-RNA	>10	>10	260/280 = 2.0 260/230 = 2.0-2.2	-	DNase digested, buffer: RNase free water
<i>cDNA Sequencing</i>	>200 ng polyA+	>10	>15	260/280 = 2.0 260/230 = 2.0-2.2	-	200 ng cDNA can be used as input DNase digested, buffer: RNase free water
<i>cDNA PCR Sequencing</i>	>10 ng polyA+ or >400 ng Total-RNA	>10	>1	260/280 = 2.0 260/230 = 2.0-2.2	-	DNase digested, buffer: RNase free water
<i>Whole Genome Sequencing</i>						
<i>Ligation Sequencing</i>	>2 $\mu\text{g}$	>50	>40	260/280 = 1.8 260/230 = 2.0-2.2	>30 kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>Native Barcoding</i>	>1 $\mu\text{g}$ per sample	>50	>20	260/280 = 1.8 260/230 = 2.0-2.2	>30 kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>Ultra Long Reads</i>	>50 $\mu\text{g}$	>750	>70	260/280 = 1.8 260/230 = 2.0-2.2	>100 kb	homogenous UHMW DNA, RNase digested, buffer: EEB (please inquire about EEB buffer)
<i>Genome Rapid</i>	>400 ng	>10	>40	260/280 = 1.8 260/230 = 2.0-2.2	>30 kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>PCR Sequencing</i>	>200 ng	>50	>4	260/280 = 1.8 260/230 = 2.0-2.2	-	clean, target-specific, buffer: EB

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<b>Oxford Nanopore Long-Read Sequencing</b>						
<i>Amplicon Sequencing</i>						
<i>Amplicons by Ligation</i>	>500 ng per sample	>10	>50	260/280 = 1.8 260/230 = 2.0-2.2	-	clean, target-specific, buffer: EB
16S	>20 ng	>10	>2	260/280 = 1.8 260/230 = 2.0-2.2	-	clean, target-specific, buffer: EB
<b>Sanger Sequencing</b>						
<i>Full Service</i>						
<i>Plasmid</i>	>500 ng	-	100 – 250	260/280 = 1.8 260/230 = 2.0-2.2	<5 kb	Buffer: 10 mM Tris/HCl or water Primer concentration 10 $\mu\text{M}$
	>800 ng	-	150 – 600	260/280 = 1.8 260/230 = 2.0-2.2	5 kb – 15 kb	Buffer: 10 mM Tris/HCl or water Primer concentration 10 $\mu\text{M}$
	>1,5 $\mu\text{g}$	-	>600	260/280 = 1.8 260/230 = 2.0-2.2	>15 kb	Buffer: 10 mM Tris/HCl or water Primer concentration 10 $\mu\text{M}$
<i>Full Service / Xpress Service</i>						
<i>PCR-product</i>	max. 10 ng	-	<2,5	260/280 = 1.8 260/230 = 2.0-2.2	<100 bp	purified PCR products Primer concentration 10 $\mu\text{M}$
	50 ng	-	< 5ng	260/280 = 1.8 260/230 = 2.0-2.2	100 bp – 1 kb	purified PCR products Primer concentration 10 $\mu\text{M}$
	400 ng	-	20 - 50	260/280 = 1.8 260/230 = 2.0-2.2	>1 kb	purified PCR products Primer concentration 10 $\mu\text{M}$

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<b>Sanger Sequencing</b>						
<i>Xpert Service</i>	-	-	-	-	-	Sample requirements as for Full Service
<i>Pre-mixed Service</i>						
	<i>Plasmid</i>	300 – 600 ng	-	-	-	<10 kb 5 pmol Primer/reaction 7,5 $\mu\text{L}$ total volume
		>700 ng	-	-	-	>10 kb 5 pmol Primer/reaction 7,5 $\mu\text{L}$ total volume
	<i>PCR-products</i>	<100 ng	-	-	-	100 bp – 1 kb 5 pmol Primer/reaction 7,5 $\mu\text{L}$ total volume
		>100 ng	-	-	-	>1 kb 5 pmol Primer/reaction 7,5 $\mu\text{L}$ total volume
<i>Ready-to-load Service</i>						
	<i>Plasmid, PCR-products</i>	-	20	-	-	Purified sequencing reaction

## Fragment Analysis

	<i>Ready-to-run</i>	-	20	-	-	<600 bp Fully prepared samples dissolved in formamide
	<i>Pre-pared</i>	-	10	-	-	<600 bp samples dissolved in formamide without length standard
<i>STR analysis</i>		50 ng	-	5	$260/280 = 1.8$ $260/230 = 2.0-2.2$	- Buffer: 10 mM Tris/HCl, low TE or water